

Available online at www.sciencedirect.com

Tetrahedron Letters 45 (2004) 371–374

Tetrahedron Letters

$BH₃$ as a protecting group for phosphonic acid: a novel method for the synthesis of dinucleoside H -phosphonate

Mamoru Shimizu, Kiyoshi Tamura, Takeshi Wada* and Kazuhiko Saigo

Department of Integrated Biosciences, Graduate School of Frontier Sciences, The University of Tokyo, Bioscience Building 702, Kashiwanoha, Kashiwa 277-8562, Japan

Received 17 September 2003; revised 20 October 2003; accepted 24 October 2003

Abstract—A $BH₃$ group is found to be an effective protecting group for phosphonic acid esters. This new phosphonic acid protecting group was applied to the synthesis of a dithymidine H-phosphonate derivative from a dithymidine boranophosphate derivative. Triarylmethyl cations were found to be effective for the deprotection of the $BH₃$ group in the dithymidine boranophosphate diester to afford the corresponding H-phosphonate derivative in excellent yield.

2003 Elsevier Ltd. All rights reserved.

H-Phosphonate DNA is regarded as a potentially useful intermediate for the synthesis of various kinds of backbone-modified nucleic acid derivatives.¹⁻¹³ However, in the methods reported so far for the synthesis of this DNA analog, undesirable side reactions occur in the chain elongation cycle and the oxidation step. $14-21$ Accordingly, only short oligomers can be synthesized in good yields by the current methods.

On the other hand, boranophosphate DNA, which is chemically and physiologically stable, has been reported as a new candidate of DNA analog.²² We recently described a new method for the synthesis of boranophosphate DNA (boranophosphotriester method).²³ It is well known that 4,4'-dimethoxytrityl (DMTr) cation reacts with boranophosphate DNA, resulting in its decomposition. Therefore, the addition of a $DMTr⁺$ scavenger is required for the detritylation reaction.^{12,23,24} In the detritylation of the boranophosphate triester 1, Et₃SiH was found to be effective as a $DMTr⁺$ scavenger for the complete suppression of the decomposition of the product. In contrast, the boranophosphate diester 3 was quickly decomposed to give the corresponding Hphosphonate diester 4, when 3 was treated with DMTr cation in the presence of $Et₃SiH$ under the same conBoranophosphate triester

Boranophosphate diester

Scheme 1. Stability of 3'-boranophosphate derivatives under detritylation conditions. (i) 3% DCA in CDCl₃–Et₃SiH (1:1, v/v).

ditions, indicating that 3 was more unstable than 1 (Scheme 1).

This result suggested that boranophosphate diester could be transformed into the corresponding H-phosphonate. In another point of view, a $BH₃$ group can be employed as a protecting group for phosphonic acids. Although a BH₃ group is frequently used as a protecting group for phosphine derivatives,²⁵ the use of a BH₃ group for the phosphonic acid protection has never been reported.

In this paper, we wish to describe the first example of the use of a $BH₃$ group as a phosphonic acid protecting group.

Keywords: protecting group; boranophosphate; H-phosphonate; phosphonic acids; oligonucleotide synthesis.

^{*} Corresponding author. Tel./fax: $+81-471-36-3612$; e-mail: [wada@](mail to: wada@) k.u-tokyo.ac.jp

^{0040-4039/\$ -} see front matter \odot 2003 Elsevier Ltd. All rights reserved. doi:10.1016/j.tetlet.2003.10.158

As a model reaction, dinucleoside boranophosphate 5, which was synthesized by the boranophosphotriester method,²³ was allowed to react with DMTr^+ , which was generated by the reaction of dimethoxytrityl methyl ether (DMTr–OMe) with trifluoroacetic acid $(TFA)^{26}$ in CDCl₃, and the reaction was monitored by ${}^{31}P$ NMR. After 10min, the broad signal of 5 (93.87 ppm) quickly disappeared, and a new signal was observed at 9.51 and 8.25 ppm (diastereomers). The P–H coupled spectrum of the reaction mixture clearly indicated the formation of dithymidine phosphonate (6) bearing a P–H bond $(J_{PH} = 719.2, 723.7 Hz)$. A plausible reaction mechanism is shown in Scheme 2.

Next, several trityl cation sources were tested for the present approach (Scheme 3).

Compound 5 was allowed to react with trityl cation reagents in the presence of TFA under various conditions (Table 1). In all cases, the broad signals of 5 (Fig. 1A) quickly disappeared, and new signals of the Hphosphonate diester 6 were observed. In addition, the broad signals of intermediates were observed around 40ppm in some cases. Although the structures of the intermediates are still not clear, these compounds were found to be readily converted to the H-phosphonate diester 6 during the aqueous work-up. For example, the quantitative formation of 6 was observed after aqueous treatment of the reaction mixture when DMTr–OMe was used (Table 1, entry 1). A similar result was obtained when DMTr–OH was employed as a trityl cation source (entry 2). In both cases, prolonged reactions (60–72 min) resulted in the formation of the P-tritylated products (28.75, 28.6 ppm, diastereomers).6 In contrast, when DMTr–Cl was employed, the formation of diastereomers of P-tritylated compounds (49.05, 48.46 ppm)²⁷ and transesterified products (6.95 ppm) were observed to some extent (entries 3 and 4). These side reactions were assumed to be catalyzed by hydro-

Scheme 2. Plausible mechanism for the transformation of *H*-phosphonate from boranophosphate diester.

Scheme 3. Structures and abbreviations of trityl reagents.

chloric acid. In order to accelerate the deprotection reaction, trimethoxytrityl (TMTr) derivatives, which generate a more stable trityl cation, were employed. As expected, the transformation reactions proceeded quickly to give the desired product 6 in excellent yields (entries 5 and 6). Although the degree of the P-tritylation upon prolonging the reaction time was slightly higher than that of the cases using DMTr derivatives, the expected product 6 was obtained quantitatively upon quenching the reaction in a short time (Fig. 1B).

Table 1. Reaction conditions^a for the transformation of boranophosphate diester 5 to H-phosphonate 6

Entry	Reagent	Additive	Ratio of compounds $(\%)^b$							
	$(5$ equiv $)$	$(5$ equiv)	Products (6)		Intermediates		P-tritylation		Transesterification ^e	
			A^c	B ^d	A	B	A	B	A	B
	DMTr-OMe		78	74	22	21				
	$DMTr-OH$		75	75	23	20				
	$DMTr-C1$		59	30	Ω	θ	20, 21 ^f	49, $21f$		
	$DMTr-Cl$	MeOH	89	72				13		15
	TMTr-OMe		95	92				х		
_n	TMTr-OH		100	79				14		
	TMTr-Cl		85	75					15	21
8	TMTr-Cl	MeOH	89	81						14
9	Pix-OMe		78	70	22	24				
10	$Pix-OH$		72	60	25	15		25		
11	$Pix-C1$		56	29	23	11		45	14	:5
12	Pix-Cl	MeOH	61	42	19			44		15

^a All the reactions were carried out with 5 (0.1 M) in the presence of TFA (3 equiv) in CDCl₃ at rt. b Estimated by ³¹P NMR analysis.

Measured during 5–10min.

 d Measured during 60–72 min.

^eMay be these side products are 5'-5' or 3'-3' dithymidine *H*-phosphonates.
 $f_{\text{Sao B of 27}}$

^fSee Ref. 27.

Figure 1. ³¹P NMR spectra of 5 (A), the reaction mixture obtained by the reaction of 5 with TMTr–OH in the presence of TFA in CDCl₃ for 1 min followed by aqueous work-up (B) , and isolated 7 in CDCl₃ (C) (P–H decoupled NMR spectra).

Scheme 4. Synthesis of O-protected TpT starting from dithymidine boranophosphate. (i) TFA, TMTr-OH/CHCl₃, (ii) Et₃SiH, ext. with CHCl₃/satd NaHCO₃, (iii) I₂/pyridine-H₂O (98:2, v/v), ext. with CHCl3/triethylammonium bicarbonate, purified by silica gel column chromatography, overall yield > 99%.

When 9-phenylxanthen-9-yl (Pix) type reagents were used as cation sources, the reaction rates were similar to those using DMTr reagents. However, a considerable amount of a mixture of the P-tritylated products was formed upon prolonging the reaction times (entries 9 and 10).

Next, the isolation of the dinucleoside H-phosphonate 6 was attempted by applying the optimized reaction conditions. Compound 5 was treated with TMTr–OH in the presence of TFA in CHCl₃ for 1 min. Although the quantitative formation of 6 was ascertained by a TLC monitoring after aqueous work-up, pure 6 was isolated only in 75% yield due to a partial decomposition of 6 during silica gel column chromatography.28;²⁹ Therefore, we attempted to isolate the product as a stable phosphate derivative, 7, after oxidation with I_2 in pyridine– H_2O (98:2, v/v).³⁰ As a result, the dinucleoside phosphate 7 was obtained in quantitative yield from boranophosphate 5 (Scheme 4 and Fig. 1C). This result clearly indicates that the deprotection of the dithymidine boranophosphate derivative did take place quantitatively.

In conclusion, we have demonstrated the first example of the use of BH_3 as a phosphonic acid protecting group. The method enabled us to synthesize *H*-phosphonate DNAs, versatile intermediates for the backbone-modified DNA analogs, by treatment of boranophosphate DNAs with a desired sequence, which can be easily prepared by the boranophosphotriester method, 23 with $Ar₃C⁺$ under mild acidic conditions. The present method will be also useful for the synthesis of a wide variety of organic compounds bearing phosphate functions as well as their analogs.

Acknowledgements

This work was supported by a Grant from a Grant-in-Aid for Scientific Research from the Ministry of Education, Culture, Sports, Science and Technology, Japan and by JSPS Research Fellowships for Young Scientists (M.S.).

References and Notes

- 1. For comprehensive reviews on H-phosphonate chemistry see: (a) Froehler, B. C. Protocols for Oligonucleotide and Analogs, Synthesis and Properties. Methods in Molecular Biology; Agrawal, S., Ed.; Humana: Totowa, 1993; Vol. 20, pp 63–80; (b) Stawinski, J. In Handbook of Organophosphorus Chemistry; Engel, R., Ed.; Dekker: New York, 1992; pp 377–434; (c) Kers, A.; Kers, I.; Kraszewski, A.; Sobkowski, M.; Szabo, T.; Thelin, R.; Stawinski, J. Nucleosides Nucleotides 1996, 15, 361.
- 2. Kume, A.; Fujii, M.; Sekine, M.; Hata, T. J. Org. Chem. 1984, 49, 2139.
- 3. Froehler, B. C. Tetrahedron Lett. 1986, 27, 5575.
- 4. Froehler, B. C.; Ng, P.; Matteucci, M. D. Nucleic Acids Res. 1988, 16, 4831.
- 5. de Vroom, E.; Dreef, C. E.; van der Elst, H.; van der Marel, G. A.; van Boom, J. H. Recl. Trav. Chim. Pays-Bas 1988, 107, 592.
- 6. de Vroom, E.; Spierenberg, M. L.; Dreef, C. E.; van der Elst, H.; van der Marel, G. A.; van Boom, J. H. Recl. Trav. Chim. Pays-Bas 1987, 106, 65.
- 7. Bollmark, M.; Zain, R.; Stawinski, J. Tetrahedron Lett. 1996, 37, 3537.
- 8. Wada, T.; Hata, T. Tetrahedron Lett. 1990, 31, 7461.
- 9. Stawinski, J.; Thelin, M. Tetrahedron Lett. 1992, 33, 7255.
- 10. Wada, T.; Sekine, M. Tetrahedron Lett. 1995, 36, 8845.
- 11. Zhang, J.; Terhorst, T.; Matteucci, M. D. Tetrahedron Lett. 1997, 38, 4957.
- 12. Higson, A. P.; Sierzchala, A.; Brummel, H.; Zhao, Z.; Caruthers, M. H. Tetrahedron Lett. 1998, 39, 3899.
- 13. Sergueev, D. S.; Shaw, B. R. J. Am. Chem. Soc. 1998, 120, 9417.
- 14. Froehler, B. C.; Matteucci, M. D. Tetrahedron Lett. 1986, 27, 469.
- 15. Strömberg, R.; Stawinski, J. Nucleic Acids Symp. Ser. 1987, 18, 185.
- 16. Gaffney, B. L.; Jones, R. A. Tetrahedron Lett. 1988, 29, 2619.
- 17. Garegg, P. J.; Lindh, J.; Regberg, T.; Stawinski, J.; Strömberg, R. Nucleosides Nucleotides 1987, 6, 655.
- 18. Garegg, P. J.; Stawinski, J.; Strömberg, R. J. Org. Chem. 1987, 52, 284.
- 19. Kuyl-Yeheskiely, E.; Spierenburg, M.; van der Elst, H.; Tromp, M.; van der Marel, G. A.; van Boom, J. H. Recl. Trav. Chim. Pays-Bas 1986, 105, 505.
- 20. Regberg, T.; Stawinski, J.; Strömberg, R. Nucleosides Nucleotides 1988, 7, 23.
- 21. Froehler, B. C.; Ng, P.; Matteucci, M. D. Nucleic Acids Res. 1986, 14, 5399.
- 22. Summers, J. S.; Shaw, B. R. Curr. Med. Chem. 2001, 8, 1147.
- 23. Wada, T.; Shimizu, M.; Oka, N.; Saigo, K. Tetrahedron Lett. 2002, 43, 4137.
- 24. Sergueeva, Z. A.; Sergueev, D. S.; Shaw, B. R. Nucleosides Nucleotides 2001, 20, 941.
- 25. Carboni, B.; Monnier, L. Tetrahedron 1999, 55, 1197.
- 26. This reaction also proceeded when dichloroacetic acid (DCA) was used in the place of TFA.
- 27. These side products are assumed to be diastereomers of Ptritylated compounds arising from a reaction at the 4'position of DMTr⁺. The structural characterization is now in progress.
- 28. Wada, T.; Kato, R.; Hata, T. J. Org. Chem. 1991, 56, 1243.
- 29. A typical procedure for the synthesis of compound 6: To a solution of 5 (34.1 mg, 0.04 mmol) and TMTr–OH (70.1 mg, 0.20 mmol) in CHCl₃ (0.4 mL), TFA (9.1 μ L) was successively added. After being stirred at rt for 1 min, $Et₃SiH$ (0.4 mL) was added and continuously stirred for another 3 min . The mixture was diluted with CHCl₃ (5 mL) and washed with saturated NaHCO₃ $(3 \times 5$ mL), and the aqueous layer was back-extracted with CHCl₃ $(2 \times 5$ mL). The organic layer and washings were combined and dried over $Na₂SO₄$, filtered, and concentrated to dryness under reduced pressure. The residue was applied to a column of silica gel (3 g). Chromatography was performed with CH_2Cl_2 , applying a gradient of MeOH (0– 4%). The fractions containing 6 were combined and concentrated to dryness under reduced pressure to give 6 $(22.1 \text{ mg}, 75%)$ as a colorless foam.
- 30. Garegg, P. J.; Lindh, I.; Regberg, T.; Stawinski, J.; Strömberg, R. Tetrahedron Lett. 1986, 27, 4051.